Des moteurs de jeux à la physique des chromosomes

Pascal Carrivain

Laboratoire de Physique Théorique de la Matière Condensée, Université Pierre et Marie Curie, 4, Place Jussieu, 75252 Paris Cedex 05, France

JURY:

Aurélien Bancaud (LAAS Toulouse)
David Bensimon (ENS Paris)
Ralf Everaers (ENS Lyon)
Jean-François Joanny (Institut Curie)
Sébastien Neukirch (Institut Jean Le Rond d'Alembert)

Directeur de thèse: Jean-Marc Victor (LPTMC Paris)









19 mars 2013



Sommaire

Présentation du laboratoire et de l'équipe M3V

Contexte

Introduction

Méthode du Pivot et du Krankshaft pour la simulation d'ADN

Modèle du "Bead spring"

SHAKE

Open Dynamics Engine

Examples

Joints mécaniques

Les différentes étapes de simulation

Manipulation de molécule unique In silico d'ADN nu

Calibration de la longueur de persistance en courbure

Calibration de la longueur de persitance en twist

Simulation In silico de fibre de chromatine

Comment compacter l'ADN?

Pinces magnétiques et fibre de chromatine

Nucleosome

Conclusion et perspectives

Sommaire

Présentation du laboratoire et de l'équipe M3V

Contexte

Introduction

Méthode du Pivot et du Krankshaft pour la simulation d'ADN

Modèle du "Bead spring

SHAKE

Open Dynamics Engine

Example

Joints mécaniques

Les différentes étapes de simulation

Manipulation de molécule unique *In silico* d'ADN n

Calibration de la longueur de persistance en courbure

Calibration de la longueur de persitance en twist

Simulation *In silico* de fibre de chromatine

Comment compacter l'ADN?

Pinces magnétiques et fibre de chromatine

Nucleosome

Conclusion et perspectives

Présentation du laboratoire et de l'équipe M3V

Modélisation Multi-échelle de la Matière Vivante

- 1. Directeur du laboratoire de Physique Théorique de la Matière Condensée : Pascal Viot
- 2. Équipe M3V:
 - Chef d'équipe : Jean-Marc Victor (Directeur de recherche au CNRS)
 - Annick Lesne (Directeur de recherche au CNRS)
 - Maria Barbi (Maitre de conférences à l'UPMC)
 - Julien Mozziconacci (Maitre de conférences à l'UPMC)
 - Laurence Signon (Chargé de recherche au CNRS et stagiaire au LPTMC)
 - ► Axel Cournac (Post-doctorant)
- 3 Anciens membres:
 - Fabien Paillusson
 - ► Hua Wong
 - Hua vvong
 - Eli Ben-Haïm

Sommaire

Présentation du laboratoire et de l'équipe M3V

Contexte

Introduction
Méthode du Pivot et du Krankshaft pour la simulation d'ADN
Modèle du "Bead spring"
SHAKE

Open Dynamics Engine

Examples

Joints mécaniques

Les différentes étapes de simulation

Manipulation de molécule unique In silico d'ADN nu

Calibration de la longueur de persistance en courbure

Calibration de la longueur de persitance en twist

Simulation *In silico* de fibre de chromatine

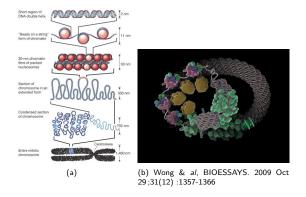
Comment compacter l'ADN?

Pinces magnétiques et fibre de chromatine

Nucleosome

Conclusion et perspectives

ADN nu sous pinces magnétiques : contexte biologique



ADN nu sous pinces magnétiques : montage expérimental

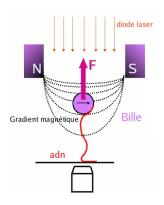


FIGURE: Vue schématique d'une expérience de pinces magnétiques.

ADN nu sous pinces magnétiques : première expérience

Direct Mechanical Measurements of the Elasticity of Single DNA Molecules by Using Magnetic Beads

Steven B. Smith, Laura Finzi,* Carlos Bustamante†

Single DNA molecules were chemically attached by one end to a glass surface and by their other end to a magnetic bead. Equilibrium positions of the beads were observed in an optical microscope while the beads were acted on by known magnetic and hydrodynamic forces. Extension versus force curves were obtained for individual DNA molecules at three different salt concentrations with forces between 10⁻¹⁴ and 10⁻¹¹ newtons. Deviations from the force curves predicted by the freely jointed chain model suggest that DNA has significant local curvature in solution. Ethidium bromide and 4',6-diamidino-2-phenylindole had little effect on the elastic response of the molecules, but their extent of intercalation was directly measured. Conversely, the effect of bend-inducing *cis*-diamminedichloroplatinum (II) was large and supports the hypothesis of natural curvature in DNA.

Propriété de courbure de l'ADN : persistance en courbure $p=50~\mathrm{nm}$ Cette expérience ne mesure pas la capacité de l'ADN à se tordre : persistance en twist t



Méthode du Pivot et du Krankshaft pour la simulation d'ADN

DNA Extension under the Action of an External Force

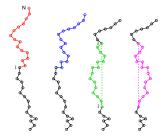
Alexander Vologodskii!

Department of Molecular and Cell Biology, University of California, Berkeley, California 94720 Received March 23, 1894; Revised Manuscript Received July 11, 1894*

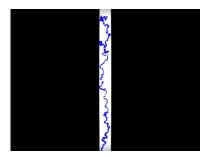
ABSTRACT. We compute the extension-force dependence for the wormlike model of a polymer chain. The dependence that we chainst differ assembled from the recreasing well-known dependence for the firely pinted chain when the attension exceeds half of the chain contour length. We used the computed results to analyze the measurement of extension of individual DNA mixedous under the excels of force valued from the control of the chain contour length. We used the computed results to analyze the measurement of the chains of individual DNA mixedous under the excels of force valued from the chain of the c

(a) Vologodskii, Macromolecules **1994**, 27, 5623-5625

Pivot Rotation $R_u(\theta)(i \rightarrow N)$ Krankshaft Rotation $R_v(\phi)(i \rightarrow j)$



(b) Vue schématique des méthodes du Pivot (de rouge à bleu) et du Krankshaft (de vert à magenta) + algorithme de Metropolis



(c) Vue schématique d'un polymère greffé dans un tube

Modèle du "Bead spring" (1)

A Brownian Dynamics Program for the Simulation of Linear and Circular DNA and Other Wormlike Chain Polyelectrolytes

Konstantin Klenin, Holger Merlitz, and Jörg Langowski Division Biophysics of Macromolecules, German Cancer Research Center, D-69120 Heidelberg, Germany

ABSTRACT For the interpretation of solution structural and dynamic data of linear and circular DNA molecules in the kb range, and for the prediction of the effect of local structural changes on the global conformation of such DNAs, we have developed an efficient and easy way to set up a program based on a second-order explicit Brownian dynamics algorithm. The DNA is modeled by a chain of rigid segments interacting through harmonic spring potentials for bending, torsion, and stretching. The electrostatics are handled using precalculated energy tables for the interactions the vene DNA segments as a function of relative orientation and distance. Hydrodynamic interactions are treated using the Rotne-Prager tensor. While maintaining acceptable precision, the simulation can be accelerated by recalculating this tensor only once in a certain number of steps.

Modèle du "Bead spring" (2)

Klonin et al BD Program for Linear and Circular DNA

FIGURE 1 Chain prometry with the segment vectors s, and the segment coordinate systems (E. g., e.) that define the relative orientation between

The total energy of a given chain conformation is given as the sum of the stretching, bending, twist, and electrostatic

The stretching energy is defined for each segment to

$$\frac{E_i^{(a)}}{k_B T} = \frac{1}{2(l_0 \delta)^2} (l_0 - \epsilon_i)^2$$

Here k_0 is the Boltzmann constant, T the temperature, l_0 the segment equilibrium length, and 5 the stiffness parameter. so that $(I_0S)^2$ is approximately equal to the variance of the segment length distribution

The bending energy is defined for each chain joint We call a joint undear if it connects segments that form a straight line at equilibrium, and bear if the angle 0" between the segments at equilibrium is nonzero. To each bent joint i we attach an auxiliary unit vector b, that is fixed in the coordinate system (f_i, g_i, e_i) , its polar coordinates being $(\theta_{i}^{+}, \varphi_{i}^{+})$. Note that this formalism is different from our first implementation (Chirico and Langowski, 1996) where a hauristic "kink notantial" was used that was given in terms of the Euler angles for rotating one segment into the next. Here the bending energy of the ith joint is:

$$\frac{E_i^{tot}}{4\sqrt{T}} = \alpha_b \beta_i^2$$

where β_i is the angle between ϵ_{i-1} , and ϵ_i for an unbent joint, or between e..., and b. for a bent joint; on in the bending rigidity parameter chosen in such a way that the Kuhn length is equal to:

$$B = I_0 \frac{1 + (\cos \beta)}{1 - (\cos \beta)},$$

where

 $\langle \cos \beta \rangle = \frac{\int_0^{\pi} \cos \beta \sin \beta \exp(-\alpha_b \beta^2) d\beta}{\int_0^{\pi} \sin \beta \exp(-\alpha_b \beta^2) d\beta}$

The twist energy is defined for each adjacent segment pair: $\frac{E_i^{(0)}}{V_i + \cdots + \frac{1}{2V_i}} = \frac{C}{2V_i + \frac{1}{2}}$

781

where C is the torsional rigidity constant and τ , is the twist angle between the (i-1)th and ith negments.

The twist angle τ_i is calculated by defining a vector p_i = s..., × s., which is normal both to s..., and s. Now, we can easily calculate the angles α_i between f_{i-1} and p_{ii} and γ_i between p, and f. Then, $\tau_i = \alpha_i + \gamma_i$ (Fig. 2). During the BD simulation we assume that at time t, $\tau(t - \Delta t) - \pi \le$ $\tau(t) \leq \tau(t - \Delta t) + \pi$ where $\tau(t - \Delta t)$ is the twist angle one simulation step ago (\Delta is chosen such that the proba-

bility that the twist angle changes by more than ± m be-The starting point for the electrostatic energy is the expression for the energy of interaction between two uniformly charged nonadjacent segments (i, j) in a 1:1 salt solution in the Debye-Hückel approximation:

$$\frac{E_0^{(s)}}{k_BT} = \frac{v^2}{k_BTD} \int d\lambda_i \int d\lambda_j \frac{\exp(-\kappa r_0)}{r_0}$$
.

The integration is done along the two segments: λ and λ are the distances from the segment beginnings, ril is the distance between the current positions at the segments to which the integration parameters λ and λ correspond: κ is the inverse of the Debye length, so that $\kappa^2 = 8\pi e^2 D k_B TD$, I is the ionic strength, e the proton charge, D the dielectric constant of water v the linear charge density which for DNA is equal to $v_{tras} = -2e/\Delta$, where $\Delta = 0.34$ nm is the distance between basenairs.

There are two problems to be solved for the Eq. 5: 1) the linear density v should be renormalized from that of DNA to a smaller value in order to ensure the correct excluded volume effects; 2) the integration should be approximated by a more simple procedure to save computation time. The renormalization of the linear density was done as in Stigter (1977). As pointed out in Schellman and Stigter (1977), the Gouy layer of immobile counterious reduces the affactive charge density by a factor of a = 0.73 for NaCl concentrations between 1 and 500 mM. Next, the Debve-Hückel approximation is a linearization of the Poisson Boltzmann equation and valid only for a very small electric potential: $\phi \ll k_a T/e$. We choose the renormalized charge



FIGURE 3 Definition of the twist angle $v_i = \alpha_i + \gamma_i$, \mathbf{p}_i is perpendicular to s. . , and s.; o, is measured between f. . , and p. . , between p. and f.

density v* in such a way that the known solution of the Debye-Hückel equation for a straight thin line with charge zmann equation for a cylinder of the DNA radius rese and the charge density qv_{cols} in the regions where $\phi \ll k_BT/e$

In order to save computation time, a tabulation of the double integral (Eq. 5) was used. For simplicity we assume that each segment has the same length L. For each segment pair (i, j) we can define a vector l_iR_i connecting the middle points of the two segments. Then the mutual position of the segments can be defined by the following four dimensionless parameters: $\rho_0 = |R_0|$

center-to-center distance in
$$L_0$$
 units, (6a $\rho_{ij} \ge 0$

 $\gamma_i = (1/\rho_i)e_i \cdot R_i$ tilt angle cosine for the 1th segment

$$\gamma_{j} = -(1/\rho_{0})e_{j}\cdot R_{0},$$
tilt angle cosine for the jth segment, (6c)

 $(e \times R_i) \cdot (e \times R_i)$ $|e \times R| |e \times R|$

twist angle cosine,
$$-1 \le \sigma_{ij} \le 1$$
 (6d)
Equation 5 can be rewritten in the form:

 $E_i^{(i)} = \alpha_i f(\rho_i, \gamma_i, \gamma_i, \sigma_i)$

where
$$\alpha_c = v^{\mu 2} l_d / k_B TD$$
 and

$$f(\rho, \gamma_1, \gamma_2, \sigma) = \int_{-12}^{12} dx_1 \int_{-12}^{12} dx_2 \frac{\exp(-\kappa |R|)}{|R|}, \quad (8)$$

 $R = \rho v_0 - x_1 v_1 + x_2 v_2$

FIGURE 3. Flactousteic naturals for the interaction harmon DNA care. ments. A Debve-Hitchel potential (I) was renormalized (3) such as to coincid at large distances with the nonlinear Possson-Boltzmann equation (2).

where v., v., and v. are unit vectors oriented in such a way that $v_0v_1 = y_1, -v_0v_2 = y_2$, and $(v_1 \times v_0) \cdot (v_2 \times v_0)$ $|\mathbf{v}_1 \times \mathbf{v}_0|\mathbf{v}_1 \times \mathbf{v}_0| = \sigma$

Volume 74 February 1998

Ricohysical Journa

In the following we need the partial derivatives of Eo. S. At first, a four-dimensional table for the values of the integral (Eo. 8) and each of its partial derivatives was constructed numerically. Then, during the simulation, a linear interpolation was used to obtain the values of $(\partial f' \partial \rho)$, (0f/0ys), (0f/0ys), (0f/0s) at particular points of the (o. ys. γ., σ) space. The table steps were chosen in such a way that the values of ρ , arccos v_{ij} arccos v_{ij} and arccos σ had constant increments. The tabulation range for v. v. and or is [-1,1]. The range of ρ , $[\rho_{min}, \rho_{max}]$, and the table size for each argument are parameters of the approximation, which we chose by the following criteria. For the minimum distance, ρ_{min} , all possible values of the electrostatic energy should be large enough (e.g., $>10 k_{\odot}T$), so that this distance is practically unreachable during the simulations. For distances $> \rho_{max}$ all possible energy values should be negligible (say, $<0.01 k_nT$). The mutual displacement of segments

corresponding to one ρ -step should not exceed the Debve length, and the same restriction is applied to the displacement of segment ends corresponding to one step in the γ_1 , y., and or dimensions. These criteria are rather "soft." in order to keen the total table size within reasonable limits (the minimum table size for $I_0 = 10 \text{ nm}$ and I = 1 M is 14 MB of memory, including all partial derivatives of Eq. 8). We should note, however, that even crude hard-core potentials for electrostatic repulsion in DNA can be applied in many cases to predict statistical properties of DNA to good precision (Vologodskii and Cozzarelli, 1995); therefore, the representation of the electrostatic potential according to For 7 and 8 is probably a good approximation So far, we neglected the fact that the segment length is

only approximately equal to I. That means that the "charged" segment does not coincide exactly with the "geometrical" segment. This seems to be a good approximation for the aveloded volume affacts, since the chain is supposed to be stiff with respect to stretching ($\delta \ll 1$). One has to be careful, however, to avoid that during the simulation parts of the chain conce each other through discontinuities has tween adjacent charged segments, if the length of their phantom recometrical counterparts is greater than Io. In order to exclude this possibility we define the R., vector for two segments (i, i) of arbitrary length in the following way:

$$R_{ij} = \frac{1}{r} (r_{ij0}^{(in)} - r_{ij0}^{(in)})$$

where $r_{(0)}^{(n)}$ is a "shifted" middle point of the segment

$$r_{i0}^{(a)} = \begin{cases} r_i + (l_0/2)\mathbf{e}_i, & \text{if } |r_i^{(a)} - r_i| \le |r_i^{(a)} - r_{i+1}|; & \text{if } |r_{i+1} - (l_0/2)\mathbf{e}_i, & \text{else}_i \end{cases}$$
(1

and $r^{(m)} = (r_1 + r_2...)/2$ is the actual middle point of the segment. This means that the geometrical segment that is used to calculate the excluded volume interaction is shifted

(a)

(b)

Modèle du "Bead spring" (3)

with its end toward the segment joint that is closest to the other segment in the interaction. Thereby, any gaps that

might appear at the joint due to stretching are automatically closed and the chains cannot cross

Since forces and torques are the partial derivatives of the energy over the system coordinates, the latter should be specified formally. For each segment i we chose the following four coordinates: the three space coordinates r, of the segment beginning and the angle φ_i of rotation of the local vector basis (f_i, g_i, e_i) around the e_i axis. For the angle coordinate the zero position is not defined. Therefore, in addition, we need to specify how to keep the q, coordinates unchanged while a displacement of the r. coordinates takes place. The following rule was used to derive forces and torques and to perform moves in the simulation procedure. If as a result of r. displacement, the e. vector has a new value e'_i and A_i is a rotation matrix, so that $e'_i = A_i e_i$, then the new value of the f, vector is $f' = A \cdot f$. In a linear chain three additional degrees of freedom ru-

are required for the last segment. Here are the expressions for the forces and torones for the ith segment obtained by differentiating Eqs. 1, 2, 4, and 7:

Stretching force

Klanin et al.

The stretching force acting on the th vertex from the (t + 1)thone is parallel to the 8th segment.

$$\frac{\mathbf{F}_{i,\text{nex}}^{(i)}}{k_BT} - \frac{z_i - l_a}{(l_a\delta)^2} \mathbf{e}_i$$

The total stretching force for the rth vertex is therefore

$$\mathbf{F}_{i}^{\omega} = -\mathbf{F}_{i-1,\text{nest}}^{\omega} + \mathbf{F}_{i,\text{nest}}^{\omega}$$

Bending force

The contribution of the energy stored in the bending angle 8, to the bending force acting on the ith vertex from the (i + 1)th one is perpendicular to the #h segment and lies in the bend plane:

$$\frac{F_{\text{born}}^{\text{blue}}}{F_0 T} = \frac{2\alpha_0 \beta_0}{s_0} p_i \times e_i \qquad (3)$$
for an unbent joint, and
$$\frac{F_{\text{born}}^{\text{blue}}}{F_0} = \frac{2\alpha_0 \beta_0}{c \sin \alpha} (e_{i-1} \times b_i) \times e_i \qquad (4)$$

for a bent joint. Here $\tilde{\mathbf{p}}_i = \mathbf{p}_i/\mathbf{p}_i$, $\mathbf{p}_i = s_{i-1} \times s_i$, so that $\tilde{\mathbf{p}}_i = -$ The torque on the #h segment induced by twisting the (i+1)th

e... × e/sin B" where B" is the angle between e... and e. (for an unbent joint, $\beta^* = \beta$.). The analogous contribution to the bending force acting on

the ith vertex from the (i-1)th one is perpendicular to the

(i - 1)th segment and also lies in the bend plane

$$\frac{\mathbf{F}_{i,pov}^{0}}{k_BT} = \frac{2\alpha_i\beta_i}{z_{i-1}}\,\mathbf{p}_i \times \mathbf{e}_{i-1} \tag{15a}$$
 for an unbent joint, and

m unbent joint, and
$$\frac{\mathbf{F}_{i,\text{per}}^{(i)}}{k_0 T} = \frac{2\alpha_i \beta_i}{z_{i-1} \sin \beta_i} [\mathbf{b}_i - (\mathbf{e}_{i-1} \cdot \mathbf{b}_i) \mathbf{e}_{i-1}] \quad (15b)$$

Note that Eqs. 14s and 15s for an unbent joint are symmetric with respect to renumbering the vertices in opposite order (one force can be obtained from the other by substituting $s_i \leftrightarrow -s_{i-1}$, $\mathbf{e}_i \leftrightarrow -\mathbf{e}_{i-1}$, $\hat{\mathbf{p}}_i \leftrightarrow -\hat{\mathbf{p}}_{i-1}$). Obviously, there is no such symmetry for a bent joint because the b, vectors are not defined in a symmetrical way. We note also that the expressions for a bent joint (14b, 15b) become those for the unbent joint upon substituting $b_i \rightarrow e_i$. The total bending force for the #th vertex is

$$\mathbf{F}^{b)} = -\mathbf{F}^{b)}_{ij-1,\text{pert}} + \mathbf{F}^{(b)}_{i,\text{pert}} + \mathbf{F}^{(b)}_{i,\text{sent}} - \mathbf{F}^{(b)}_{ij+1,\text{pert}} \quad (16)$$

Bending torque

for a bent joint

BD Program for Linear and Circular DNA

The torque on segment i induced by bending is for a bent

$$\frac{T_i^{(b)}}{k_B T} - \frac{2\alpha_b \beta_i}{\sin \beta_i} \mathbf{e}_{i-1} \cdot (\mathbf{e}_i \times \mathbf{b}_i) \qquad (1$$

For an unbent joint, this torque is equal to zero.

Twisting force

The force on the th vertex from the (t + 1)th one induced by mutual twisting of the (t - 1)th and th segments by an angle 7, is perpendicular to the plane of the 8th bend:

$$\frac{\mathbf{F}_{t_0}^{(i)}}{k_0T} = \frac{C}{k_0T\,l_0} \frac{\tau_{F_1-1}}{p_i} (1-\epsilon_{i-1} \cdot \epsilon_i) \hat{\mathbf{p}}_i \tag{18}$$
The symmetric expression for the twisting force acting on

the #h vertex from the (1 - 1)th one is $\frac{\mathbf{F}_{i,per}^{ij}}{\mathbf{k}.T} = -\frac{C}{\mathbf{k}_{z}TL}\frac{\tau_{z}s_{i}}{\mathbf{p}_{i}}(1 - \mathbf{e}_{i-1} \cdot \mathbf{e}_{i})\mathbf{p}_{i}$ (19)

$$\frac{k_0T}{k_0T} = \frac{1}{k_0T}\frac{T_0}{p_0}(1 - \epsilon_{i-1} \cdot \epsilon_i)p_i$$
The total twisting force for the rith vectex is then
$$F_i^{ij} = -F_{ij}^{(i)} \cdot \epsilon_{i+1} + F_{ij}^{(i)} = +F_{ij}^{(i)} - F_{ij}^{(i)} \cdot \epsilon_{i+1}$$

(a)

segment by an angle 7,..., with respect to the #h one is

$$\frac{T_{k_{\text{min}}}^{0}}{k_{\text{B}}T} = \frac{C}{k_{\text{B}}T I_{0}} \tau_{i+1} \qquad (21)$$

The total twisting torque for the #th segment is then $T^{0} = -T^{0}_{0}, \dots + T^{0}_{m}$

Electrostatic force

783

The contribution of the electrostatic interaction between segments (and (to the force acting on the 8h vertex is

$$\frac{\mathbf{F}_{0,1}^{(a)}}{k_BT} = \frac{1}{-k_BT} \frac{\partial \mathbf{E}_0^{(a)}}{\partial \mathbf{r}},$$

$$= \alpha_i \left(\frac{\partial f_i}{\partial \rho_j} \frac{\partial \rho_i}{\partial \mathbf{r}_i} + \frac{\partial f_i}{\partial \gamma_j} \frac{\partial \gamma_i}{\partial \mathbf{r}_i} + \frac{\partial f_i}{\partial \gamma_j} \frac{\partial \gamma_i}{\partial \mathbf{r}_i} + \frac{\partial f_i}{\partial \alpha_j} \frac{\partial \sigma_i}{\partial \mathbf{r}_i} \right)$$
where $f_{ij} = f(\rho_i, \gamma_{ij}, \gamma_{ij}, \sigma_{ij})$. An analogous relationship is valid for the force $F^{(2)}$ scenar on the $ii + 1$ lth vertex from

segment j. We assume that $\mathbf{F}_{k1}^{(i)} = \mathbf{F}_{k2}^{(i)} = 0$ when i = i or $i = j \pm 1$. The partial derivatives of f (Eq. 8) are tabulated as described above. The expressions for the derivatives of ρ_{ii} , γ_{ii} , γ_{ij} , and σ_{ij} with respect to r_i and r_{i+1} are given below, where the following auxiliary notations are used: $\hat{R}_{..} = R_{..}/\rho_{..}$ is a unit vector in the R_{a} direction; α_{av} and β_{av} are the coefficients in the expression for the "shifted" mid dle-point of a segment:

$$r_{ij}^{(n)} = \alpha_{ij} r_i + \beta_{ij} r_{i+1},$$
 so that (see Eq. 11):

 $\alpha_{in} = (1/2)(1 \pm (1 - L/s))$

 $\beta_{co} = (1/2)(1 \mp (1 - L/\xi))$ The upper variant corresponds to the condition (a) in Eq. 11.

the lower variant corresponds to the condition (b) The partial derivatives are thus: $\frac{\partial \rho_0}{\partial t} = -\frac{1}{t} \alpha_{av} \hat{R}_a \mp \frac{1}{2} \frac{\gamma_0}{\theta} e_i$

$$\frac{\partial p_{ij}}{\partial r_{i+1}} = -\frac{1}{l_0} \beta_{ij} \hat{R}_{ij} = \frac{1}{2} \frac{\gamma_{ij}}{z_i} e_i \qquad (24) \qquad D_{ij} = D_{ij} \frac{3\sigma}{4r_{ij}} \left[I + \frac{r_{ij} \otimes r_{ij}}{r_{ij}^2} + \frac{2\sigma^2}{3r_{ij}^2} \left(I - 3 \frac{r_{ij} \otimes r_{ij}}{r_{ij}^2} \right) \right] \qquad (24) \qquad if \quad r_{ij} \approx 2\sigma, \quad i \neq i$$

$$\begin{array}{lll} & & & & \\ & & & \\ \frac{\partial \gamma_t}{\partial r_t} - \frac{1}{\rho_0} \left[\frac{1}{b_0} a_0 \rho_0 \left[\frac{\partial b_t}{\partial r_t} + \frac{\partial \gamma_{t+1}}{\partial r_t} \right] & & \\ & & & \\ \frac{\partial \gamma_t}{\partial r_t} - \frac{1}{\rho_0} \left[\frac{1}{b_0} a_0 \rho_0 + \frac{(r_t \cdot \mathbf{e}_t)}{2r_t} + \rho_0 \frac{\partial \rho_0}{\partial r_t} \right] & & \\ & & & \\ & & & \\ & & & \\ \end{array} \right] & & \text{where } r_0 = r_1 - r_2 \cdot r_2 = |r_0|, D_0 = h_0 \text{Tr} \text{Graps}, \eta \text{ is the water} \\ & & \\ & & \\ & & \\ & & \\ \end{array}$$

$$\frac{\partial \gamma_{ji}}{\partial r_{i+1}} = \frac{1}{\rho_{ij}} \left[\frac{1}{l_0} \beta_{ijj} e_j \mp \frac{(e_i \cdot e_j)}{2t_i} e_i - \gamma_{ij} \frac{\partial \rho_{ij}}{\partial r_{i+1}} \right] \qquad (28)$$

Biophysical Journal

$$\begin{split} \frac{\partial \sigma_{ij}}{\partial r_{i}} &= \sqrt{1 - \sigma_{ij}^{2}} \left[\frac{s_{j}\gamma_{ij}}{|\vec{R}_{ij} \times s_{j}|^{2}\rho_{ij}} \left[\frac{1}{l^{2}}\alpha_{iij}(\vec{R}_{ij} \times s_{j}) \right. \\ &\left. + \frac{1}{2} \frac{(\vec{R}_{ij} \cdot (s_{j} \times s_{i}))}{s_{i}^{2}} \sigma_{ij} - \left[\frac{\vec{R}_{ij} \times s_{ij}}{|\vec{R}_{ij} \times s_{ij}|^{2}} \left(1 - \frac{\alpha_{iij}s_{j}\gamma_{ij}}{l_{i}\rho_{ij}} \right) \right], \end{split} \tag{29}$$

Volume 74 February 1998

$$\frac{\partial \sigma_{ij}}{\partial r_{i+1}} = \sqrt{1 - \sigma_{ij}^2} \left\{ \frac{s_i \gamma_{ij}}{|\hat{R}_{ij} \times s_j|^2 \rho_{ij}} \left[\frac{1}{l_0^2} \beta_{ij}(\hat{R} \times s_i) \right. \\
\left. + \frac{1}{2} \frac{(\hat{R}_{ij} \cdot (s_j \times z_i))}{s_i^2} e_i \right] - \frac{\hat{R}_{ij} \times s_i}{|\hat{R}_{ij} \times s_i|^2} \left(1 + \frac{\beta_{ij} z_i \gamma_{ij}}{k \rho_{ij}} \right) \right\}; (30)$$

 $\sqrt{1-\sigma_{ij}^{\alpha}}$ is taken when $R_{ij} \cdot (s_i \times s_j) > 0$, and its negative value otherwise. The total electrostatic force acting on the #h vectex is

$$\mathbf{F}_{i}^{(s)} = \sum_{i}^{N-1} (\mathbf{F}_{0,i}^{(s)} + \mathbf{F}_{0-1y,2}^{(s)})$$
 (3)

The boundary conditions for the force expressions, Eos. 12-31 are different for linear and circular chains. For a linear chain all parameters with indexes out of range do not exist and can be formally set to zero. The allowed ranges are $0 \le i \le N-1$ for $\mathbf{F}_{i,\text{new}}^{(b)}$, $\mathbf{I}_{j,\text{new}}^{(b)}$, $1 \le i \le N-1$ for $\mathbf{F}_{j,\text{new}}^{(b)}$, $\mathbf{F}_{j,$ conditions are in effect, such that all indices have to be taken modulo N.

Hydrodynamic interactions

In order to model hydrodynamic interactions defined between unherical objects a head with radius a was attached to each chain vertex. With N'=N+1 beads for a linear and N' = N beads for a circular chain, we describe the hydrodynamic interaction between beads i and j by a 3×3

$$_{ij} = D_{0} \frac{\delta a}{4r_{ij}} \left[I + \frac{r_{ij} \otimes r_{ij}}{r_{ij}^{2}} + \frac{2a^{\prime}}{3r_{ij}^{2}} \left(I - 3 \frac{r_{ij} \otimes r_{ij}}{r_{ij}^{2}} \right) \right],$$

if $r_{ij} \approx 2a$, $i \neq j$; (32a)

if
$$r_{ij} \le 2a$$
, $i \ne j$; (32b)
 $D_{ii} = D_{ij}J$ (32c)

$$q=[r_{ij}],D_0=k_{\rm H}T/6\pi\eta\sigma,\eta$$
 is the water 3×3 matrix, and $r\otimes r$ denotes a matrix

with the components $(r_{\alpha}r_{\beta})$; α , $\beta = x, y, z$. The bead radim a is connected with the DNA hydrodynamic radius r_{in} , in the following way. Let us consider two

(b)

SHAKE

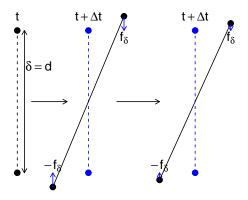


FIGURE: Vue schématique d'un algorithme type SHAKE.

Edberg & al, J. Chem. Phys. 84 (12), 15 June 1986
Forester & al, Journal of Computational Chemistry, Vol. 19, No. 1, 102-111 (1998)
Hess & al, Journal of Computational Chemistry, Vol. 18, No. 12, 1463-1472 (1997)
Ryckaert & al, Journal of Computational. Physics 23, 321-341 (1977)

Sommaire

Présentation du laboratoire et de l'équipe M3V

Contexte

Introduction

Méthode du Pivot et du Krankshaft pour la simulation d'A

Modèle du "Bead spring"

SHAKE

Open Dynamics Engine

Examples

Joints mécaniques

Les différentes étapes de simulation

Manipulation de molécule unique In silico d'ADN nu

Calibration de la longueur de persistance en courbure

Calibration de la longueur de persitance en twist

Simulation *In silico* de fibre de chromatine

Comment compacter l'ADN?

Pinces magnétiques et fibre de chromatine

Nucleosome

Conclusion et perspectives

Examples





(a) Starcraft 2

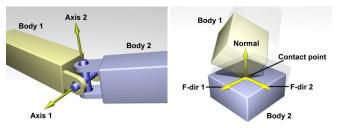
(b) Crysis 3

Joints mécaniques

ODE is Copyright 2001-2004 Russell L. Smith. All rights reserved.



- (a) Vue schématique d'un joint "Ball-in- (b) Vue schématique d'un joint "Hinge" Socket"



- (c) Vue schématique d'un joint de Cardan
- (d) Vue schématique d'un joint de contact

Joints mécaniques : formulation mathématique

Équation de la dynamique d'un système de corps rigides

$$S = \{\mathbf{r}_{1}, \mathbf{q}_{1} \dots \mathbf{r}_{N}, \mathbf{q}_{N}\}^{T}$$

$$V = \{\mathbf{v}_{1}, \omega_{1} \dots \mathbf{v}_{N}, \omega_{N}\}^{T}$$

$$\mathcal{F}_{e} = \{\mathbf{f}_{1}, \Gamma_{1} \dots \mathbf{f}_{N}, \Gamma_{N}\}^{T}$$

$$\mathcal{M} = \{\mathbf{m}_{1}, \mathcal{I}_{1} \dots \mathbf{m}_{N}, \mathcal{I}_{N}\}$$

$$\dot{\mathcal{L}} = \mathcal{F}_{e} \text{ avec } \mathcal{L} = \mathcal{MV}$$

- ▶ Contrainte holonomique : $\delta(S, t) = 0$ (joints mécaniques)
- lacktriangle Contrainte cinématique : $\dot{oldsymbol{\delta}}={oldsymbol{\mathcal{JV}}}=0$
- lacktriangle Torseur dynamique des contraintes ${\cal F}_c={\cal J}^{\sf T} {\pmb \lambda}$, avec ${\pmb \lambda}$ à calculer!
- ▶ Le torseur des contraintes ne travaille pas $\mathcal{F}_c^T \mathcal{V} = 0!$
- $\qquad \qquad \bullet \ \left[\boldsymbol{\mathcal{J}} \boldsymbol{\mathcal{M}}^{-1} \boldsymbol{\mathcal{J}}^{T} + \tfrac{k_{\mathsf{cfm}}}{\Delta t} \right] \boldsymbol{\lambda} = \tfrac{k_{\mathsf{erp}}}{\Delta t^{2}} \boldsymbol{\delta} \boldsymbol{\mathcal{J}} \left[\tfrac{\boldsymbol{\mathcal{V}}}{\Delta t} + \boldsymbol{\mathcal{M}}^{-1} \left(\boldsymbol{\mathcal{F}}_{e} \dot{\boldsymbol{\mathcal{M}}} \boldsymbol{\mathcal{V}} \right) \right]$
- $ightharpoonup k_{
 m erp}$ et $k_{
 m cfm}$ servent à contrôler la stabilité du système



Les différentes étapes de simulation

- 1. N cylindres de longueur $I=3.34~\mathrm{nm}$ et de rayon effectif $r_{\mathrm{eff}}=r_c+l_D$ (r_c représente le rayon cristallographique de l'ADN et l_D la longueur de Debye de la solution en sel) connectés par des joints "Ball-in-Socket"
- 2. Couples de courbure et de twist $\Gamma_{b+t}=g_b\mathbf{t}_1\times\mathbf{t}_2+\frac{g_t\phi}{1+\cos\theta}\left(\mathbf{t}_1+\mathbf{t}_2\right)$

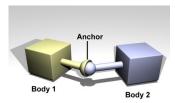


FIGURE: Vue schématique d'un joint "Ball-in-Socket"

- 3. Dynamique de Langevin-Euler locale de corps rigides $\mathcal{G} = -\Sigma \mathcal{L} + \Xi \dot{\mathcal{W}}$
- 4. ODE calcule les contraintes λ
- 5. Le système évolue du temps t au temps $t+\Delta t$ avec une intégration semi-implicite

Dynamique de Langevin-Euler avec un thermostat global (1)

Berendsen & al, J. Chem. Phys. 81 (8), 15 October 1984 Bussi & al, The Journal of Chemical Physics 126, 014101 (2007) Bussi & al, Computer Physics Communications 179 (2008) 2629

- ► Thermostat global versus thermostat local
- ▶ Hamiltonien du système : $H(S, V) = T(v) + R(\omega) + U(S)$
- ▶ Distribution dans l'ensemble canonique : $P(S, L) dSdL \propto e^{-\beta H(S, bsmcV)}$
- ▶ Vitesse de thermalisation de l'énergie cinétique :

$$dH = \sum_{i=1}^{6N} \left[\left(\frac{\Sigma_{ii}^{\star}}{\beta} - \frac{\Sigma_{ii}^{\star}}{\mathcal{M}_{ii}^{\star}} \mathcal{L}_{i}^{\star 2} \right) dt + \sqrt{\frac{2\Sigma_{ii}^{\star} \mathcal{L}_{i}^{\star 2}}{\beta \mathcal{M}_{ii}}} dW_{i} \right]$$

$$= (\langle E \rangle - 2T) \Sigma_{T}^{\star} dt + (\langle E \rangle - 2R) \Sigma_{R}^{\star} dt + 2\sqrt{\frac{\Sigma_{T}^{\star} T + \Sigma_{R}^{\star} R}{\beta}} dW$$

- "Disturbance" $(\dot{\mathcal{L}} \mathcal{F}_e)^T \mathcal{M}^{-1} (\dot{\mathcal{L}} \mathcal{F}_e) = \mathcal{G}^T \mathcal{M}^{-1} \mathcal{G}$
- $\qquad \qquad \text{Dynamique de Langevin-Euler globale}: \ \widetilde{\mathcal{G}} = \left[\Sigma^{\star} \left[\frac{\langle E \rangle}{E} \left(1 \frac{1}{2\beta \langle E \rangle} \right) 1 \right] + \sqrt{\frac{\Sigma^{\star}}{\beta E}} \dot{\mathcal{W}} \right] \mathcal{L}$
- $ightharpoonup \widetilde{\mathcal{G}}$ n'intervient pas dans le calcul du torseur des contraintes \mathcal{F}_c



Dynamique de Langevin-Euler avec un thermostat global (2)

Diffusion de la distance bout à bout d'une molécule d'ADN de longueur L:

$$\left\langle \left(\mathsf{R}\left(t + \tau \right) - \mathsf{R}\left(t \right) \right)^{2} \right\rangle = \left\langle \mathsf{R}^{2}\left(t + \tau \right) \right\rangle - 2 \left\langle \mathsf{R}^{T}\left(t + \tau \right) \mathsf{R}\left(t \right) \right\rangle + \left\langle \mathsf{R}^{2}\left(t \right) \right\rangle \tag{1}$$

Cas limite $\tau \gg \tau_{\text{deccorelation}}(\mathbf{R})$:

$$\langle (\mathbf{R}(t+\tau) - \mathbf{R}(t))^2 \rangle \simeq \langle \mathbf{R}^2(t+\tau) \rangle + \langle \mathbf{R}^2(t) \rangle = 4Lp$$
 (2)

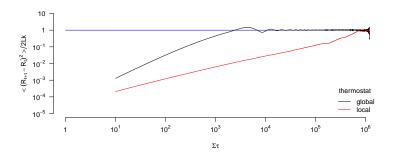


FIGURE: Diffusion de la distance bout à bout normalisée de la molécule d'ADN (en ordonnée) en fonction du temps adimensionné de simulation (en abscisse).

Sommaire

Présentation du laboratoire et de l'équipe M3V

Contexte

Introduction

Méthode du Pivot et du Krankshaft pour la simulation d'ADN

Modèle du "Bead spring

SHAKE

Open Dynamics Engine

Example

Joints mécaniques

Les différentes étapes de simulation

Manipulation de molécule unique In silico d'ADN nu

Calibration de la longueur de persistance en courbure

Calibration de la longueur de persitance en twist

Simulation In silico de fibre de chromatine

Comment compacter l'ADN?

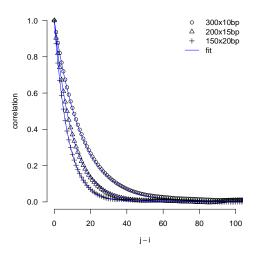
Pinces magnétiques et fibre de chromatine

Nucleosome

Conclusion et perspectives

Calibration de la longueur de persistance en courbure (1)

Corrélation tangentielle le long de la molécule d'ADN et validation des moyennes angulaires. Paramètres de la simulation $p=50~\mathrm{nm},\ t=95~\mathrm{nm},\ et\ L_{ADN}=1~\mu\mathrm{m}.$



 $\text{Figure: Corrélation tangentielle } \mathbf{t}_{i}^{T}\mathbf{t}_{j} = \exp{-|j-i|} l/p \text{ (en ordonnée) le long de la molécule d'ADN (en abscisse)}.$

Calibration de la longueur de persistance en courbure (2)

Molécule d'ADN sous force de traction. Paramètres de la simulation $p=50~\mathrm{nm},\,t=95~\mathrm{nm},\,$ et $L_{\mathrm{ADN}}=1~\mathrm{\mu m}.$

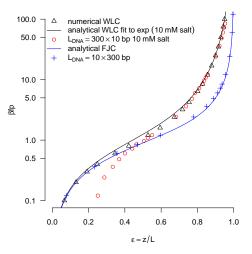


FIGURE: L'extension relative de la molécule d'ADN (en abscisse) est montrée en fonction de la force de traction adimensionnée (en ordonnée). Les résultats de simulations sont confrontés aux résultats expérimentaux de Bustamante, 1992, Siggia 1995.

Vue schématique de plectonèmes à nombre de tours fixés obtenus avec ODE

Paramètres de la simulation $\rho=50~\mathrm{nm},~t=95~\mathrm{nm},~L_{\mathrm{ADN}}=1~\mu\mathrm{m},$ avec une force de traction $f=0.74~\mathrm{pN}$ et un nombre de tour de bille magnétique n=15.

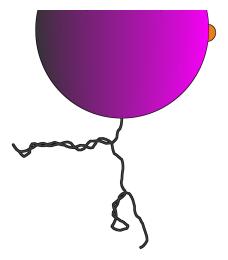


FIGURE: Vue schématique de plectonèmes à nombres de tours fixés (vidéo).

Calibration de la persistance en twist : simulation à nombre de tours fixé

Extension relative de la molécule d'ADN à nombre de tours de bille magnétique fixé. Paramètres des simulations $p=50~\mathrm{nm},$ $t=95~\mathrm{nm},$ $r_{\mathrm{eff}}\simeq2~\mathrm{nm}$ et $L_{\mathrm{ADN}}=1~\mathrm{\mu m}.$

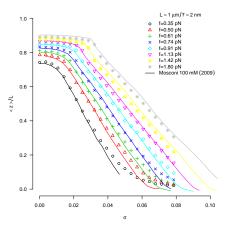


FIGURE: L'extension relative de la molécule d'ADN (en ordonnée) est montrée en fonction du nombre de tours de bille magnétique (en abscisse) pour une différentes forces de traction. Les symboles représentent les résultats de simulations et les lignes pleines représentent les résultats expérimentaux (Mosconi & al, PRL 102, 078301 (2009) solution de 100 mM de sel).

Vue schématique de plectonèmes à couple fixé obtenus avec ODE

Paramètres de la simulation $p=50~\mathrm{nm},~t=95~\mathrm{nm},~L_{ADN}=1~\mu\mathrm{m},$ avec une force de traction $f=0.74~\mathrm{pN}$ et un couple sur la bille magnétique $\Gamma_n=15~\mathrm{pN}\cdot\mathrm{nm}.$

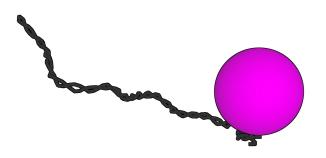
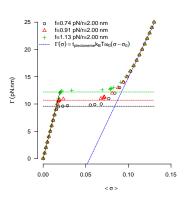


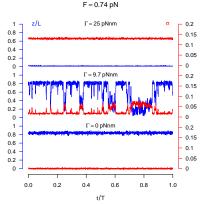
FIGURE: Vue schématique de plectonèmes à couple fixé (vidéo).

Calibration de la persitance en twist : simulation à couple fixé

Nombre de tours de la bille magnétique en fonction du couple appliqué. Paramètres des simulations $p=50~\mathrm{nm},~t=95~\mathrm{nm},~r_{\mathrm{eff}}\simeq 2~\mathrm{nm}$ et $L_{\mathrm{ADN}}=1~\mathrm{\mu m}.$



(a) L'overtwist (en abscisse) est montré en fonction du couple appliqué sur la bille magnétique. Les lignes en pointillées sont des estimations analytiques des couples de flambage (Mosconi & al PRL 102, 078301 (2009)). Le couple est déduit des expériences à nombre de tours fixés (Zhang & al, PRE 77, 031916 (2008)).



(b) L'overtwist (en rouge) et l'extension relative (en bleu) de la molécule d'ADN sont montrés (en ordonnée) en fonction du temps de simulation (en abscisse) et ce pour trois couples $\Gamma=0,9.7,25~\mathrm{pN}\cdot\mathrm{nm}$ et une force de traction $f=0.74~\mathrm{pN}.$

Sommaire

Présentation du laboratoire et de l'équipe M3V

Contexte

Introduction

Méthode du Pivot et du Krankshaft pour la simulation d'ADN

Modèle du "Bead spring

SHAKE

Open Dynamics Engine

Examples

Joints mécaniques

Les différentes étapes de simulation

Manipulation de molécule unique *In silico* d'ADN nu

Calibration de la longueur de persistance en courbure

Calibration de la longueur de persitance en twist

Simulation In silico de fibre de chromatine

Comment compacter l'ADN?

Pinces magnétiques et fibre de chromatine

Nucleosome

Conclusion et perspectives

Comment compacter l'ADN?







(b) Configuration d'une molécule d'ADN surenoulée



(c) 7 tétrasomes



(d) 4 nucléosomes

Pinces magnétiques et fibre de chromatine

Structural plasticity of single chromatin fibers revealed by torsional manipulation

Aurélien Bancaud¹, Natalia Conde e Silva², Maria Barbi³, Gaudeline Wagner¹, Jean-François Allemand⁴, Julien Mozziconacci³, Christophe Lavelle²,³, Vincent Croquette⁴, Jean-Marc Victor³, Ariel Prunell² & Jean-Louis Viovy¹

Nucleosome Chiral Transition under Positive Torsional Stress in Single Chromatin Fibers

Aurélien Bancaud, ^{1,7,6} Gaudeline Wagner, ^{1,7} Natalia Conde e Silva, ² Christophe Lavelle, ^{2,3,4} Hua Wong, ³ Julien Mozziconacci, ³ Maria Barbi, ³ Andrei Sivolob, ⁶ Eric Le Cam, ⁴ Liliane Mouawad, ⁶ Jean-Louis Viovy, ¹ Jean-Marc Viotor, ³* and Ariel Prunell^{2*}.



Figure 1 Schematic of the experiment. A single nucleosome array (~7.5 kbp), sandwiched between two naked DNA spacers (~600 bp each), is linked to a coated surface and to a magnetic bead. A pair of magnets placed above this molecule exerts controlled torsional and extensional constraints⁸.

Modèle gros-grain de nucléosome

Analyse des modes normaux (J. Mozziconacci) et structuration mécanique (P. Carrivain et H. Wong)

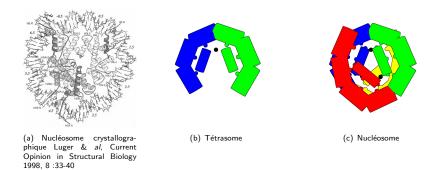


FIGURE: Vue schématique du modèle gros-grain de nucléosome construit à partir de la structure crystallographique. L'ADN nucléosomal est enroulé à gauche. Les dimères H2A-H2B sont représentés en rouge et jaune tandis que les dimères H3-H4 sont représentés en bleu et vert. Les sphères noires représentent les "Four-Helix-Bundle".

Nucléosome en traction

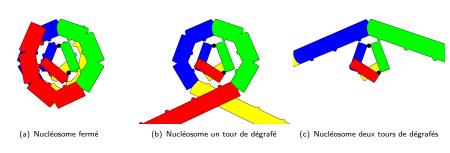


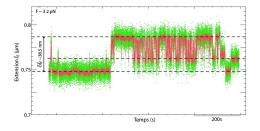
FIGURE: Vue schémtatique des trois états du nucléosome sous force de traction, modèle communèment admis dans la littérature.

- ▶ Kruithof & al, Biophysical Journal Volume 96 May 2009 37083715
- ► Ettig & al, Biophysical Journal Volume 101 October 2011 19992008
- **.**..



Nucléosome en traction : données d'E. Praly

Avec D. Bensimon, V. Croquette, F. Ding, J. Mozziconacci, E. Praly et J-M. Victor



 $FIGURE\ 14.5-Suivi\ de\ l'extension\ d'un substrat mononucléosomal insensible à la torsion, à force constante \\ \sim 3.2\ pN, en fonction\ du temps. Les données brutes correspondent aux points verts; un moyennage de ces données, sur <math>0.25\ s,$ est donné en rouge.

FIGURE: Figure tirée de la thèse d'É. Praly (Étude du mécanisme d'action des facteurs de remodelage de la chromatine, à l'échelle de la molécule unique). Nous pouvons distinguer trois états d'extension associés à deux sauts de 12 nm puis 24 nm.

Nucléosome en traction : dégraffage symétrique de l'ADN

Avec D. Bensimon, V. Croquette, F. Ding, J. Mozziconacci, E. Praly et J-M. Victor

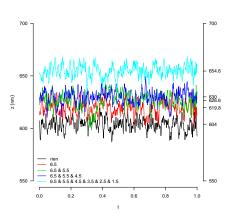


FIGURE: Schéma de dégraffage symétrique de l'ADN



Nucléosome en traction : dégraffage assymétrique de l'octamère (1)

Avec D. Bensimon, V. Croquette, F. Ding, J. Mozziconacci, E. Praly et J-M. Victor

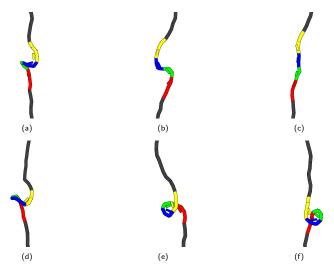


FIGURE: Ces images illustrent les différentes conformations accessibles au nucléosome pour un dégraffage assymétrique de l'octamère. La simulation est réalisée pour une force de traction de $3.2~\mathrm{pN}$ et un couple nul $\Gamma=0$.

Nucléosome en traction : dégraffage assymétrique de l'octamère (2)

Avec D. Bensimon, V. Croquette, F. Ding, J. Mozziconacci, E. Praly et J-M. Victor

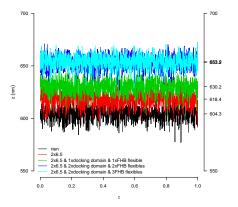


FIGURE: Schéma de dégraffage assymétrique de l'octamère. Deux sauts en extension de 12 nm et 23 nm.

Nucléosome en traction : modélisation dynamique

Avec D. Bensimon, V. Croquette, F. Ding, J. Mozziconacci, E. Praly et J-M. Victor

Modélisation des interactions avec des potentiels de Morse

Nucléosome sous une force de traction $f=3.2~\mathrm{pN}$ et un couple de $\Gamma=0~\mathrm{pN}\cdot\mathrm{nm}$ Calcul de structure pour obtenir les interactions au sein du nucléosome

SHLs	$D(k_BT)$	$a \text{ (nm}^{-1})$	r_e (nm)
±6.5	2	3.46	0.20
± 5.5	10	0.80	0.87
± 4.5	15	0.51	1.35
± 3.5	20	0.78	0.89
± 2.5	15	0.85	0.81
± 1.5	15	0.62	1.12
±0.5	20	0.85	0.81

(a) Paramètres pour les SHLs.

Interaction dimère-dimère	$D(k_BT)$	$a \left(\text{nm}^{-1} \right)$	r_e (nm)
Docking Domain	5	0.69	1.00
Docking Domain	5	0.69	1.00
Four-Helix-Bundle	15	0.39	1.76
Four-Helix-Bundle	20	0.40	1.70
Four-Helix-Bundle	15	0.39	1.76

⁽b) Paramètres pour les DDs et FHBs.

FIGURE: Tableau récapitulatif des paramètres des potentiels de Morse utilisés pour reproduire les SHL, FHB et DD.

Nucléosome en traction et torsion

Avec D. Bensimon, V. Croquette, F. Ding, J. Mozziconacci, E. Praly et J-M. Victor

Réversome : nucléosome sous une force de traction $f=3.2~\mathrm{pN}$ et un couple de $\Gamma=25~\mathrm{pN}\cdot\mathrm{nm}$

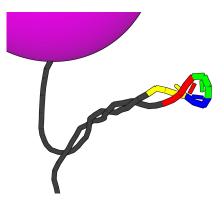
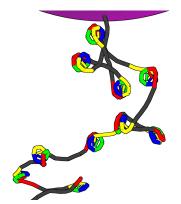


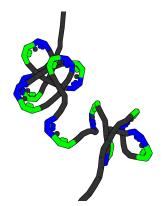
FIGURE: Vue schématique d'un réversome au milieu de plectonèmes : nucléosome sous une force de traction $f=1~\mathrm{pN}$ et un couple de $\Gamma=10~\mathrm{pN}\cdot\mathrm{nm}$.

Réversome à l'intérieur d'une fibre de chromatine

Assemblages de nucléosomes et tétrasomes sous une force de traction de $f=1~\mathrm{pN}$ et un couple $\Gamma=10~\mathrm{pN}\cdot\mathrm{nm}$

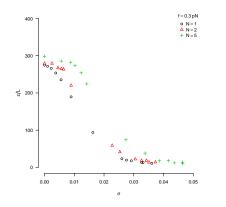


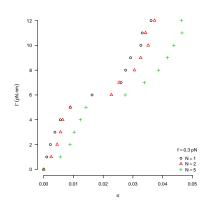
(a) Réversome à l'intérieur d'une fibre de chromatine



(b) Réversome à l'intérieur d'une "fibre" d'Archaea

Fibre de chromatine sous force de traction et couple appliqué sur la bille magnétique





- (a) L'extension de la fibre de chromatine (avec 1, 2, 5 nucléosomes) est montrée en fonction du nombre de tours pour une force de traction $f=0.3~\mathrm{pN}$.
- (b) Le couple appliqué sur la bille magnétique est montré en fonction du nombre de tours de la bille magnétique. Le couple de flambage est de l'ordre de $\Gamma_{\text{D}}=6~\mathrm{pN}\cdot\mathrm{nm}$

FIGURE:



Sommaire

Présentation du laboratoire et de l'équipe M3V

Contexte

Introduction

Méthode du Pivot et du Krankshaft pour la simulation d'ADN

Modèle du "Bead spring

SHAKE

Open Dynamics Engine

Example

Joints mécaniques

Les différentes étapes de simulation

Manipulation de molécule unique In silico d'ADN n

Calibration de la longueur de persistance en courbure

Calibration de la longueur de persitance en twist

Simulation *In silico* de fibre de chromatine

Comment compacter l'ADN?

Pinces magnétiques et fibre de chromatine

Nucleosome

Conclusion et perspectives

Conclusion et perspectives

- 1. Nouvel outil pour la simulation de macromolécules : moteurs de jeux
- 2. Images des structures complexes telles que les fibres de chromatine sous contraintes
- 3. Finir le calcul de structure du nucléosome
- 4. Mesures de la diffusion des plectonèmes
- Simulation du "crosslink" des expériences de HiC (Avec A. Cournac, J. Mozziconacci et J-M. Victor)



 Simulation de fibre de chromatine inhomogène : les cinq couleurs de la chromatine chez la drosophile (Avec G. Cavalli et C. Vaillant)



 Simulation des complexes de protéines "polycomb" et du noyau de drosophile (Avec G. Cavalli et C. Vaillant)